Oxford Molecular Haematology Laboratory, Level 4 John Radcliffe Hospital Headington Oxford OX3 9DU

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Oxford Genetics Laboratories

Solid Tumour Molecular Diagnostics Request Form

| Patient Detai | ıls | | | | | | | |
|--|--------|--|-----------------------------------|----------|------------------------|--------------------------|---------------------|----------------------|
| NHS No: | | | | Se | x: | | | |
| Surname: | | | | Ad | ldress: | | | |
| Forename: | | | | | | | | |
| Date of Birth: | | | | Po | stcode: | | | |
| Hospital: | | | | Но | spital No: | | | |
| Referral hospital pathology case number and block number/letter: | | | | | | | | |
| Ethnicity: | | | | + | IS or private? | | | |
| Date of biopsy: | | | | Da | te of request: | | | |
| | | | | | | | | |
| Referrer Det | alis | | | - | :1/DI | l | | |
| Clinician: | | | Email/Phone: Invoice address | | | | | |
| Reporting Address: | | | | | fferent: | | | |
| Address: | | | | II ai | iierent: | | | |
| Tumour Details | | | | | | | | |
| Sample type: | | | | | Tumour histology: | | | |
| Site of tumour in | this | | | | Primary site of tumour | | | |
| biopsy: | | | | | (e.g. lung, co | lon): | | |
| Tumour content | across | | >30%: Please send as Scrolls (see | | | | ease send as slide- | |
| cut surface: | | | sample requireme | nts f | or details) | | | sections (see sample |
| | | | | | | | requirem | ents for details) |
| Estimated percentage of neoplastic tissue in area for extraction: | | | | | | | | |
| (scrolls: across cut surface, sections: within marked region of H and E) | | | | | | | | |
| Test Required (Targets as Defined by Test Directory) | | | | | | | | |
| M1.1 Colorectal Small Variant (DNA) panel | | | | | | | | |
| M1.5 Colorectal Cancer – MLH1 Promotor Methylation Testing | | | | | | | | |
| M3.6 Breast Cancer – PIK3CA | | | | | | | | |
| M4.1 NSCLC Lung Cancer Small Variant (DNA) Panel | | | | | Include M4.2 * | | | |
| (For M4.2 see additional Sample Requirements) | | | | | | | | |
| M7.1 Melanoma Small Variant (DNA) Panel | | | | | | | | |
| M7.2 Melanoma <u>BRAF Only</u> | | | | | | | | |
| M215.5 Endometrial Cancer – <i>POLE</i> | | | | | | | | |
| M215.2 Endometrial Cancer – <i>MLH1</i> Promotor Methylation Testing | | | | | | | | |
| M8.1 Gastrointestinal Stromal Tumour Small Variant (DNA) Panel | | | | | | | | |
| M9.1 Thyroid Papillary Carcinoma Small Variant (DNA) Panel | | | | | | | | |
| M10.1 Thyroid Follicular Carcinoma Small Variant (DNA) Panel | | | | | | | | |
| M11.1 Poorly Differentiated Anaplastic Thyroid Carcinoma Small Variant (DNA) Panel | | | | | | | | |
| DNA-based MSI Testing | | | | | | Discuss prior to sending | | |
| Neurological Tumours - DNA Methylation Array | | | | | | | | |
| M21.1 Astrocytoma (Adult) Small Variant (DNA) Panel | | | | | | | | |
| | | | | | | | | |

Modified: 19/04/2024

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| M27.1 Glioblastoma (Adult) Small Variant (DNA) Panel | | | |
|--|--|--|--|
| M28.1 Glioma (Adult) Small Variant (DNA) Panel | | | |
| M33.3 Meningioma (Adult) Small Variant (DNA) Panel | | | |
| M36.12 Pilocytic Astrocytoma (Adult) Small Variant (DNA) Panel | | | |
| M35.1 Oligodendroglioma (Adult) Small Variant (DNA) Panel | | | |
| M184.1 Glioma (Paediatric) Small Variant (DNA) Panel | | | |
| M185.1 High Grade Glioma (Paediatric) Small Variant (DNA) Panel | | | |
| For other Test Directory indications please contact the laboratory directly to discuss prior to referral | | | |

Sample Requirements

One of:

- Samples with >30% neoplastic nuclei across the cut surface: ~50 µm FFPE tissue provided as scrolled sections cut at 6-8µm in an appropriately labelled 1.5ml microcentrifuge ("Eppendorf") tube.
- Samples with <30% neoplastic nuclei across the cut surface: Ten serial 5 micron sections (or five if marked tumour area >2cm2 or up to 15 for very small samples) mounted on microscope slides. Multiple sections can be placed on a single slide. Microtome blade and water-bath should be cleaned thoroughly before cutting sections to avoid cross-contamination and false positive results.
 - Please also include an H&E stained section from the same block with the tumour boundary marked. Percentage of neoplastic content in the marked area should be >30%
- Cytological material should be processed as FFPE tissue and sections provided as above, or the maximum available material (smears, touch preps etc) provided on slides
- * In addition to DNA based testing we are able to arrange M4.2 from a partner laboratory; if this is required please send an additional 5x scrolled sections in an appropriately labelled microcentrifuge ("Eppendorf") tube.

Labelling Standards

Please label slides with a minimum of the patient's surname and block identifier. Scrolled sections should be provided in similarly labelled microcentrifuge tubes. A minimum of 2 identifiers must be provided or the sample cannot be accepted for testing.

| Address for Sending | | | | | | |
|---|---|--|--|--|--|--|
| Please send slides to: | Molecular Haematology, Level 4, John Radcliffe Hospital, Oxford OX3 9DU. Tel: 01865 572769. E-mail: molecularhaem.oxfordgenetics@ouh.nhs.uk | | | | | |
| If you prefer to send FFPE blocks, these will need to be cut and the sections marked by a Histopathologist prior to testing, therefore please send blocks to: | Molecular Diagnostic Requests, Department of Cellular Pathology, Level 1, John Radcliffe Hospital, Headington, Oxford OX3 9DU. Tel: 01865 220499/572771. E-mail: molpath@ouh.nhs.uk | | | | | |

In submitting this sample the clinician confirms that consent has been obtained for testing and storage. Anonymised stored samples may be used for quality control procedures including validation of new genetic tests.

Information for Patients

Blood samples can be arranged via your GP or the phlebotomy clinic of your local hospital. This form must accompany the

Following receipt of the sample, laboratory staff are unable to provide information on samples and test results directly to patients or their relatives. Such enquiries should be directed to the referring clinical teams or the GP.

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